

Studying the Presence and Distribution
of Pollutants and Other Water Quality
Parameters along the Quinnipiac River
2022-2025

Courtney McGinnis Ph.D.
August 10, 2025



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Courtney McGinnis Ph.D.
Interim Associate Dean for Academic Affairs and Strategic
Initiatives of the College of Arts and Sciences
Department of Biological Sciences
Environmental Science and Studies Program
Frank H. Netter Department of Medical Sciences

Contact me @ Courtney.mcginnis@qu.edu



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Relevance

More CT towns are finding PFAS in their water supplies

Smaller water systems could face huge expenses to treat the 'forever chemicals,' or they could look for new sources of water

by Andrew Brown
August 27, 2023 @ 5:00 am

LOCAL NEWS

Class action lawsuit accuses major Connecticut water suppliers of supplying water contaminated with PFAS

Three lead plaintiffs represent hundreds of thousands of Connecticut Water and Aquiferion customers.



Author: Sakina Hossain
Published: 8:57 PM EDT October 16, 2023
Updated: 9:22 PM EDT October 23, 2023

- Per- and polyfluoroalkyl substances (PFAS) are an emerging pollutant
- 2018 (Federal –AFFF ban)
- Oct 1, 2021 (CT State)
- March 14, 2023: drinking water standard for 6 PFAS compounds

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PFBS

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PFBA

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PFOS

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PFOA

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GenX

CT's public water systems may soon need to treat for PFAS

The 'forever chemicals' have already been found in water supplies across Connecticut, and a new federal regulation could establish an enforceable limit

by Andrew Brown
April 9, 2023 @ 5:00 am



The Metropolitan District Commission's Riverside Water Treatment facility in New Hartford, the MDC is one of the state's largest public water systems. All public water systems in Connecticut may soon need to test for PFAS chemicals, and they will need to filter out these chemicals if they are found. www.ct.gov/mdc

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Per/polyfluoroalkyl substances : The forever chemicals

1930 Initial production of Polytetrafluoroethylene (PTFE)

1970 Emerging health concerns

2000s Cessation of long chain PFAS production by 3M

Electrochemical Fluorination → Telomerization process

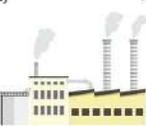
Rapid development and increasing applications → Stockholm Conventions lists PFAS as Persistent Organic Pollutants

New generation of PFAS with uncertain impacts

1930 Firefighting foams



1930s Textile industries



1940s Pesticides



1950s Protective coatings PFOA



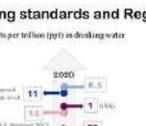
1960s Firefighting foams PFOS



1970s Non-stick cookware



1980s Paints



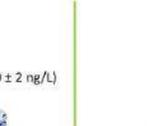
1930s Manufacturing stain and water-resistant products PFOS



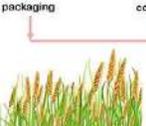
1940s Fast-food packaging



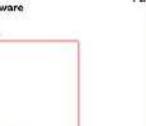
1950s Crops



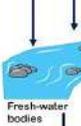
1960s Agriculture



1970s Human-exposure



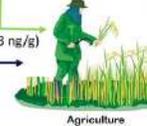
1930s Fresh-water bodies (0.1-33ppt)



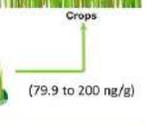
1940s Waste-water treatment plant (20 ± 2 ng/L)



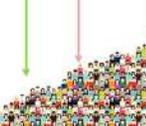
1950s (1.08-7.53 ng/g)



1960s (79.9 to 200 ng/g)

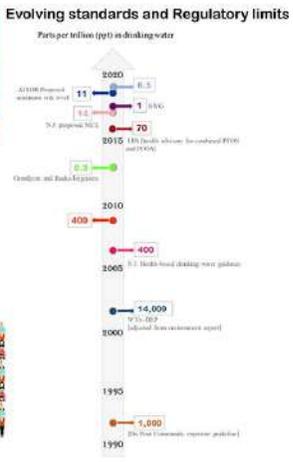


1970s (1.04-16.66 ng/ml)



Evolving standards and Regulatory limits

Parts per trillion (ppt) in drinking water



<https://www.sciencedirect.com/science/article/pii/S0960852421011494>

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Investigate the impact of per- and polyfluoroalkyl substances (PFAS) on human health



- My research students and I are currently investigating potential mechanism of actions (MOA) for several PFAS compounds.
 - Selection of the compound
 - Screening of nuclear hormone receptors and other genes associated with cholesterol transport
 - Cardiovascular disease
- This work is currently being carried out in *C. elegans*, but will transition to *D. rerio*.
- Students learn a variety of molecular lab techniques
- Students learn a variety of soft skills too!

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The Quinnipiac River



<https://www.thequinnipiacriver.com/map>

- 38-mile river that runs from New Britain-Farmington area
- The river then meanders south through Meriden and Wallingford, and ultimately drains into New Haven Harbor and Long Island Sound.
- By the Numbers:
 - 38 miles long
 - 165 square mile watershed
 - Travels through 14 municipalities
 - 913 acres of tidal marsh
 - 20 tributaries

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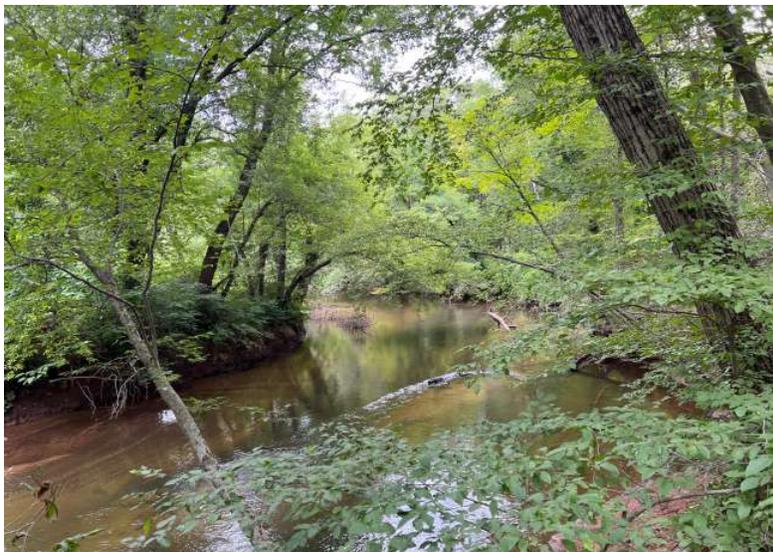
Research Overview of 2022-2023

- Focus on analyzing plasticizers, phthalates, and environmental pollutants via GC-MS
- Thirteen (13) sampling sites along the Quinnipiac River
- Sampled nine (9) times from May-August (Summer only)
- Total and Fecal Coliform Analysis



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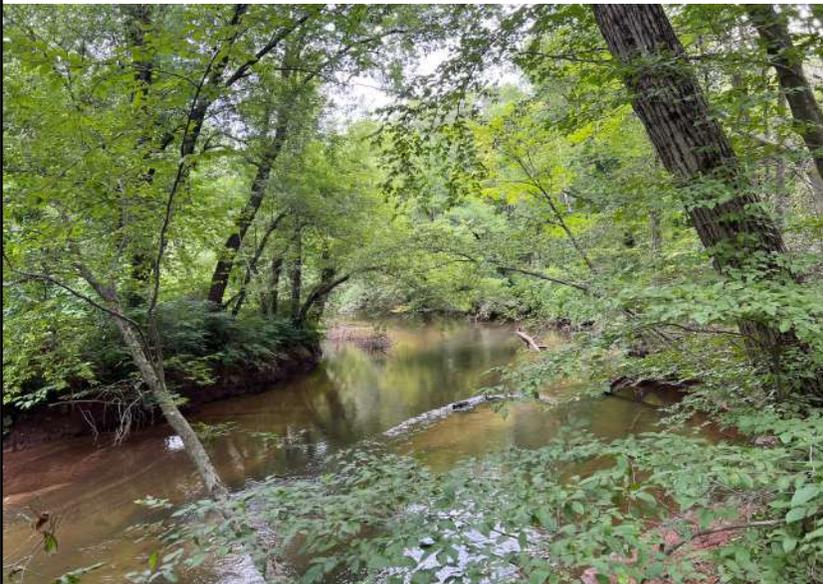
Research Overview 2023-2024



- Focus on seasonally sampling and analyzing plasticizers, phthalates, and environmental pollutants via GC-MS
- Thirteen (13-14) sampling sites along the Quinnipiac River, depending on if the drainpipe is running
- Seasonally sampling, completed 6 sampling dates so far; goal is 12 sampling dates.
 - (3x) April- June
 - (2x) July- September
 - (3x) October-December
 - (2x) January-March
- Total and Fecal Coliform Analysis

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Research Overview 2024-2025

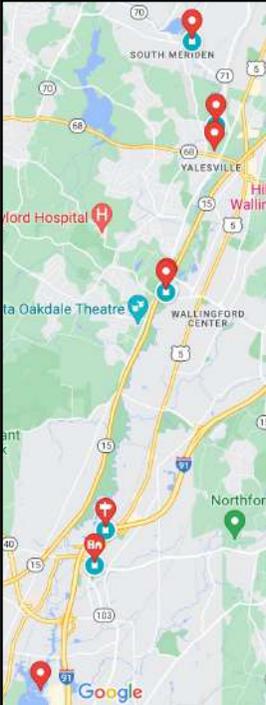


- Focus on seasonally sampling and analyzing plasticizers, phthalates, and environmental pollutants via GC-MS
- Thirteen (13-14) sampling sites along the Quinnipiac River, depending on if the drainpipe at Toelle's road is running
- Seasonally sampling
 - (3x) April- June
 - (3x) July- September
 - (3x) October-December
 - (3x) January -March
- Total and Fecal Coliform Analysis



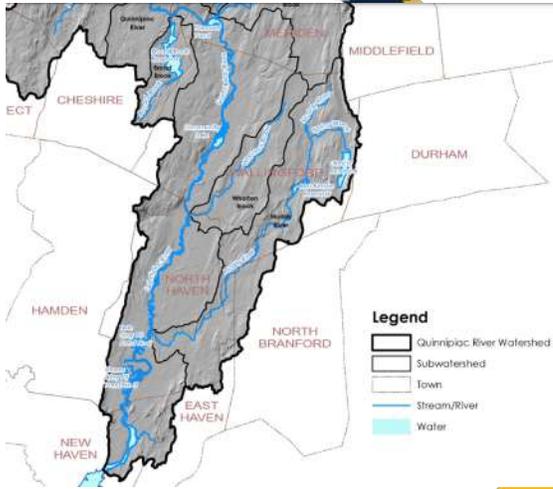
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Map of Sampling Sites



Sampling locations:

- Hanover Pond, Meriden
- Pragemann Park, Wallingford
- Fishway Above & Below, Wallingford
- Hall Ave, Wallingford
- Toelles Road, Wallingford
- Drain Pipe (not always running), Wallingford
- Valley Service, North Haven
- Quinnipiac River State Park Entrance, North Haven
- River St. Tavern, North Haven
- DEEP Boat Launch, North Haven
- Sacket Point Rd, North Haven
- Best Buy, North Haven
- Target, North Haven



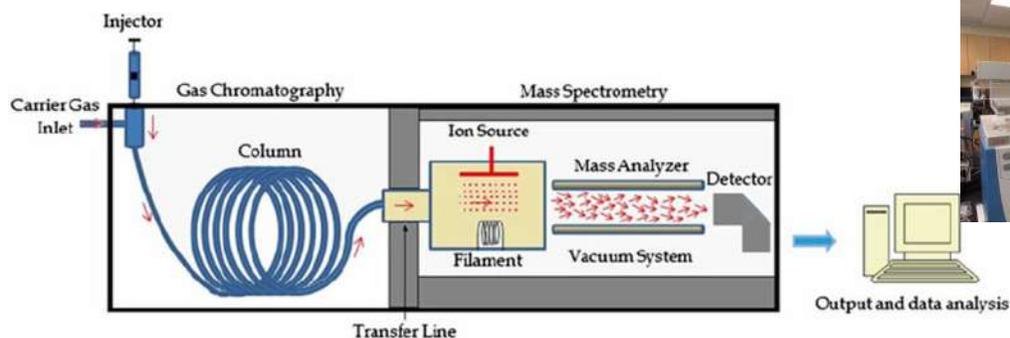
https://portal.ct.gov/-/media/DEEP/water/watershed_management/wm_plans/quinnipiac/quinnipiacwbappendicespdf.pdf



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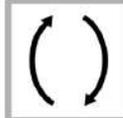
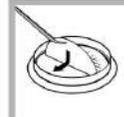
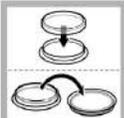
Method of Analysis

- A GC/MS was used to analyze the collected samples
- The GC (Gas Chromatographer) is used to separate the sample into smaller components so the MS (Mass Spectrometer) can measure the mass-to-charge ratio of the organic compounds and determine the presence of compounds



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Membrane filtration test procedure

			
1. Invert one m-ColiBlue24 broth ampule 2 to 3 times. Open the ampule. Lift the lid of a petri dish and carefully pour the contents equally on the absorbent pad.	2. Set up the membrane filtration apparatus. Use a sterile forceps to put a membrane filter in the assembly. Make sure that the grid side is up.	3. Invert the sample or the diluted sample for 30 seconds (25 times) to make sure that the sample is mixed well.	4. Pour or use a pipet to add the sample into the funnel. If the volume is less than 20 mL, add 10 mL of sterile buffered dilution water to the funnel.
			
5. Apply the vacuum until the funnel is empty. Stop the vacuum.	6. Rinse the funnel with 20 to 30 mL of sterile buffered dilution water. Apply the vacuum. Rinse the funnel two more times.	7. Stop the vacuum when the funnel is empty. Remove the funnel from the filter assembly. Use sterile forceps to lift the membrane filter.	8. Put the membrane filter on the absorbent pad. Let the membrane filter bend and fall equally across the absorbent pad to make sure that air bubbles are not caught below the filter.
			
9. Put the lid on the petri dish and invert the petri dish.	10. Incubate the inverted petri dish at $35 \pm 0.5^\circ\text{C}$ ($95 \pm 0.9^\circ\text{F}$) for 24 hours.	11. Remove the petri dish from the incubator. Use a 10 to 15x microscope to count the number of bacteria colonies on the membrane filter. Refer to interpret and report the coliform results on page 5.	

Method of Analysis

Total and Fecal Coliform mColiBlue

- 24-hour incubation @ 35 degrees Celsius
- EPA Method 1604
- Water Quality Criteria
 - “all recreational uses” The geometric mean density of indicator bacteria must be less than 126 colonies/100 ml and the single sample maximum is limited to 576 colonies/100 ml to comply with CT’s indicator bacteria criteria. (US EPA, 2008)

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Summer 2022 Findings

- Ph range throughout all sites, for all collections
 - 7.23-8.79
- Temp range
 - 18.5-26.8 degrees Celsius
 - May 17th - July 19th
- Conductivity range
 - 1500uS- 428uS
 - Target-Fishway/Whiteway
- July 12th large fish death, similar to previous years
- The drain pipe was running on two of the sampling days
- No Significant Findings on the GC-MS



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Summer 2022 Findings

- Beginning with the June collections all water samples we diluted 1:100 prior to analysis for total and fecal coliform.
- Samples prior to that were scored as either “Confluent growth with coliforms” or “Too Numerous To Count”
- **All samples exceed the EPA's STV value for recreational water (cfu/100mL)**

Table 4. Recommended 2012 RWQC.

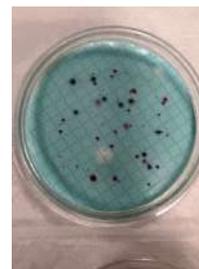
Criteria Elements	Estimated Illness Rate (NGI): 36 per 1,000 primary contact recreators		OR	Estimated Illness Rate (NGI): 32 per 1,000 primary contact recreators	
	Magnitude			Magnitude	
Indicator	GM (cfu/100 mL) ^a	STV (cfu/100 mL) ^a		GM (cfu/100 mL) ^a	STV (cfu/100 mL) ^a
Enterococci – marine and fresh	35	130		30	110
OR					
<i>E. coli</i> – fresh	126	410		100	320

Duration and Frequency: The waterbody GM should not be greater than the selected GM magnitude in any 30-day interval. There should not be greater than a ten percent excursion frequency of the selected STV magnitude in the same 30-day interval.

^a EPA recommends using EPA Method 1600 (U.S. EPA, 2002a) to measure culturable enterococci, or another equivalent method that measures culturable enterococci and using EPA Method 1603 (U.S. EPA, 2002b) to measure culturable *E. coli*, or any other equivalent method that measures culturable *E. coli*.



No dilution



1:100

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Spring & Summer 2023 Findings

7/25/23	pH	Conductivity (uS)	8/14/23	pH	Conductivity
Target	7.75	765	Target	7.52	22mS
Best Buy	7.39	731	Best Buy	7.36	15.88mS
Sacket Point Rd	7.67	435	Sacket Point Rd	7.78	946uS
DEEP	7.7	451	DEEP	7.59	3.47mS
River St. Tavern	7.36	438	River St. Tavern	8.19	465uS
Valley Service	7.51	430	Valley Service	7.8	481uS
QRSP Entrance	7.59	425	QRSP Entrance	7.87	427uS
Toeles Road	7.63	429	Drain Pipe	7.93	516uS
Hall Ave	7.68	420	Toeles Road	7.93	516uS
Fishway Below Dam	7.68	371	Hall Ave	7.95	482uS
Fishway Above Dam	7.69	421	Fishway Below Dam	8.08	483uS
Prageman Park	7.56	350	Fishway Above Dam	7.99	483uS
Hanover Pond	8.38	375	Prageman Park	8.07	465uS
			Hanover Pond -(40)	8.94	458uS

- GC-MS findings, hydrocarbons present within the Quinnipiac River Marsh (Sacket point, DEEP, Best buy and Target)
- Heavy, and frequent rain events made sampling challenging this year
 - More runoff, high levels
- 36.5% of samples (May-Sept) did not have any E.coli present, only total coliform



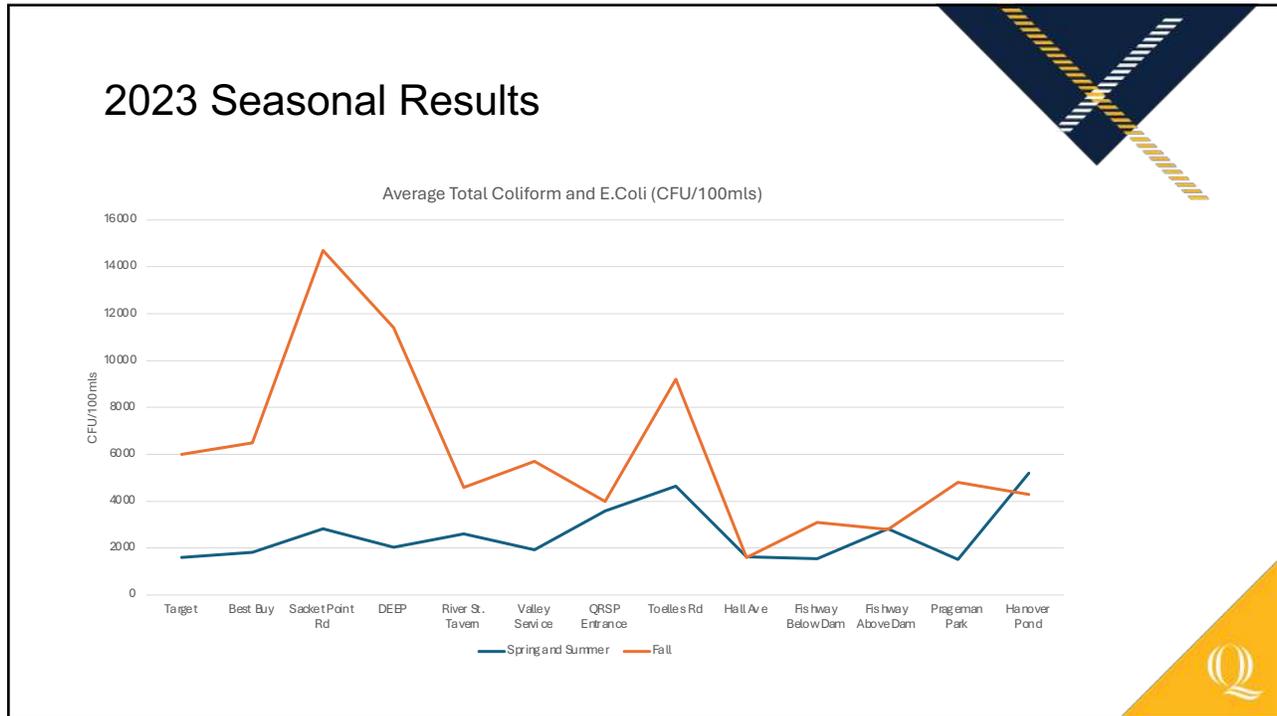
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Fall 2023 Findings

- The rain was really challenging for our Early Fall sampling, but we managed to sample 3x in this quarter
- 15% of the samples did not have E.coli present, only total coliform
- Average across all sites: 6,053CFU/100ml
 - Range 1600 (Hall Ave)- 14,700CFU (Sacket Point Rd)



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2023 Seasonal Results

Sampling Season	pH range	Temp range (°C)	Conductivity range (µS)	CFUs/100mL
1	7.44-8.2	18.4-19.3	440-1900	400-7500
2	7.36-8.94	21.6-26.4	350-1588	100-11800
3	7.16-10.37	5.5-16.7	362-2010	1600-24700
4	7.8-10.77	5.8-6.4	315-2103	5300-11200

1=April-June; 2=July-Sept; 3=Oct-Dec; 4=Jan-Mar)

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Spring & Summer 2024 Findings

June 10, 2024	Sample #	pH	Conductivity (uS)	Temp
Target	127	7.96	1129	19.6
Best Buy	126	7.63	5410	19.0
Sacket Point Rd	128	8.23	474	19.3
DEEP	125	7.98	479	19.4
River St. Tavern	124	7.47	440	19.2
Valley Service	123	7.95	424	18.0
QRSP Entrance	122	8.40	442	17.7
Toelles Road	121	8.45	455	17.8
Hall Ave	120	8.51	437	17.8
Fishway Below Dam	119	8.61	438	17.7
Fishway Above Dam	118	8.72	439	17.5
Prageman Park	117	8.69	420	17.6
Hanover Pond	116	9.17	401	18.8

- GC-MS findings, hydrocarbons present within the Quinnipiac River Marsh
 - 93% of the samples had no evidence of phthalates or plasticizers
- 30.0% of samples (May-Sept) did not have fecal coliform present, only total coliform



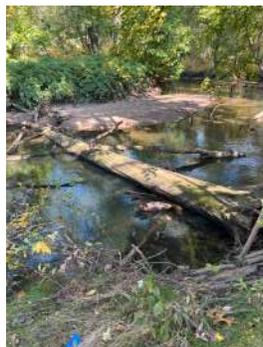
Kate Skidmore, sampling the QU River



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Fall 2024 Findings

- The river was very low October 11th
- 36.3% of the samples from this quarter did not have fecal coliform present.
- Average across all sites: 6,381CFU/100ml (+328 CFU from 2023)
 - Range 1300 (Fishway Below Dam)- 17,700CFU (River Street Tavern)

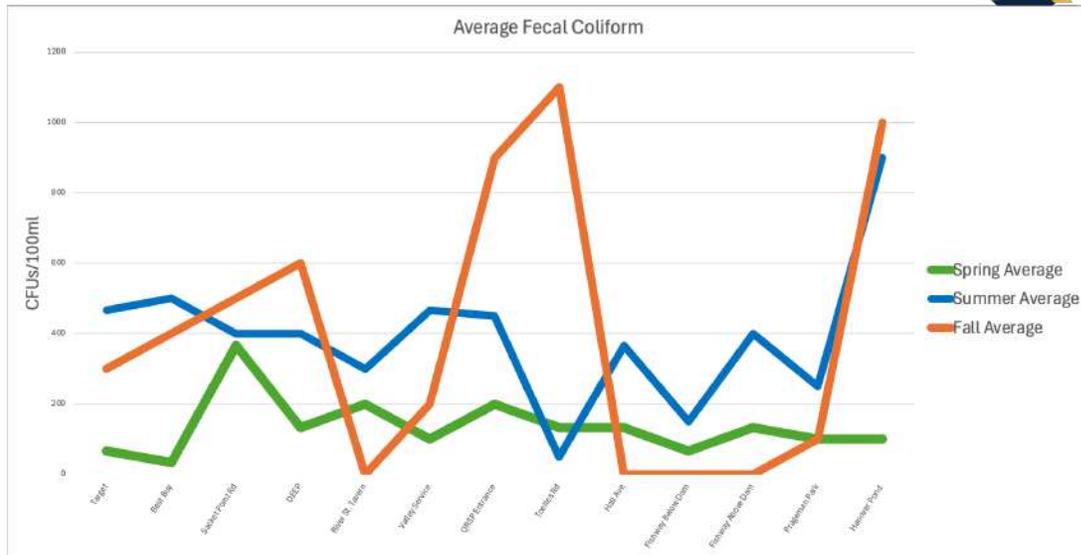


Dr. Encarnacao, sampling the QU River



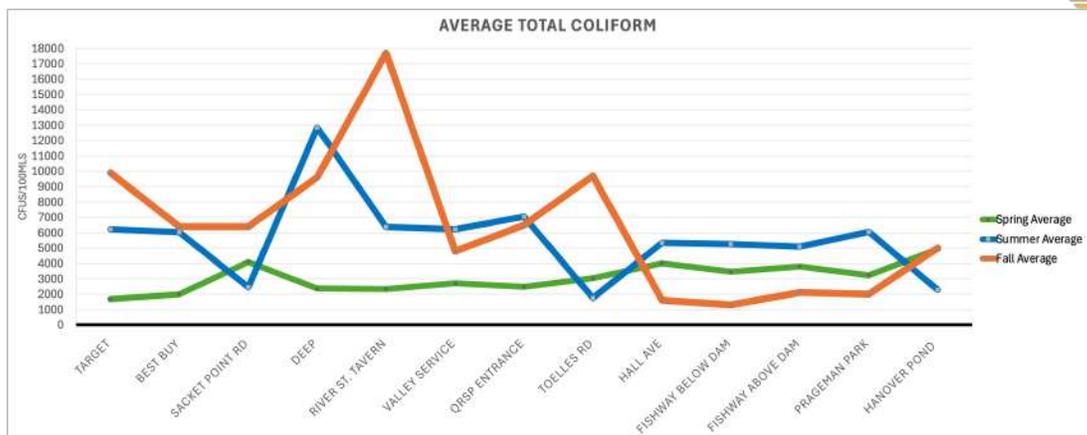
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2024 Seasonal Results



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2024 Seasonal Findings



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April 2024 Cleanup Event: QU Big Event



YOU GOT THIS, KID!
Leadership Foundation



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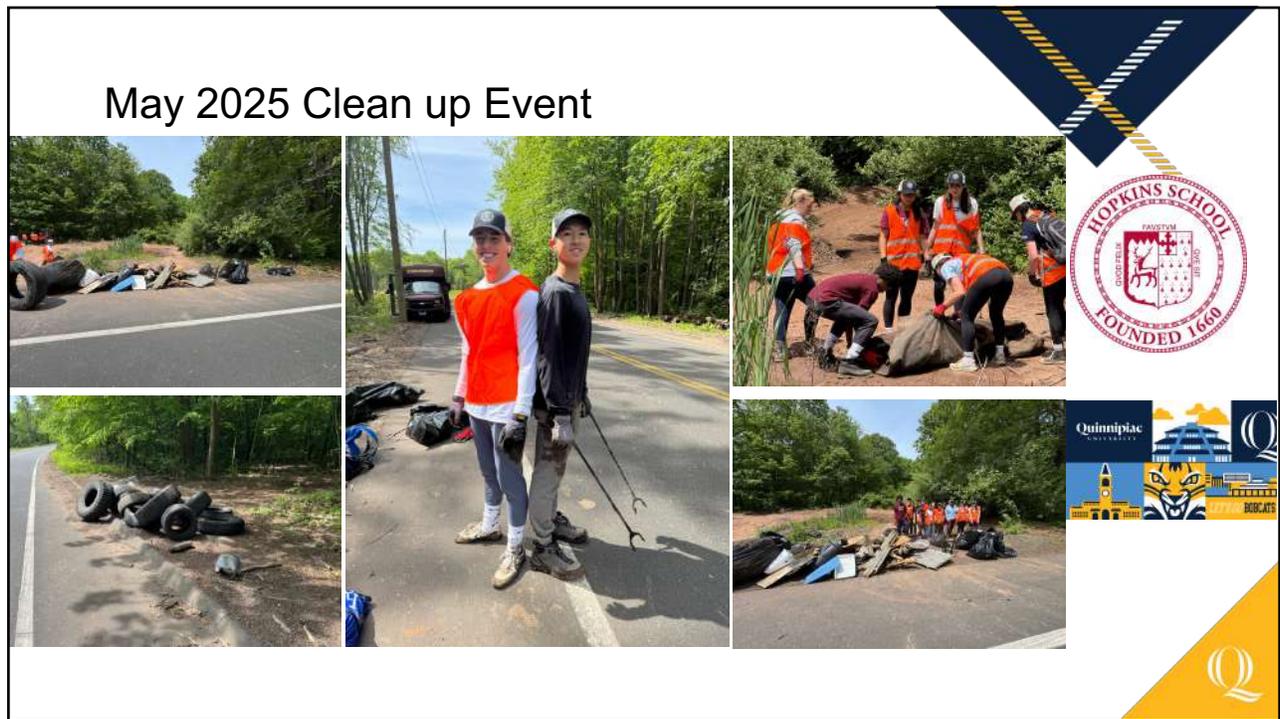
May 2024 Cleanup Events



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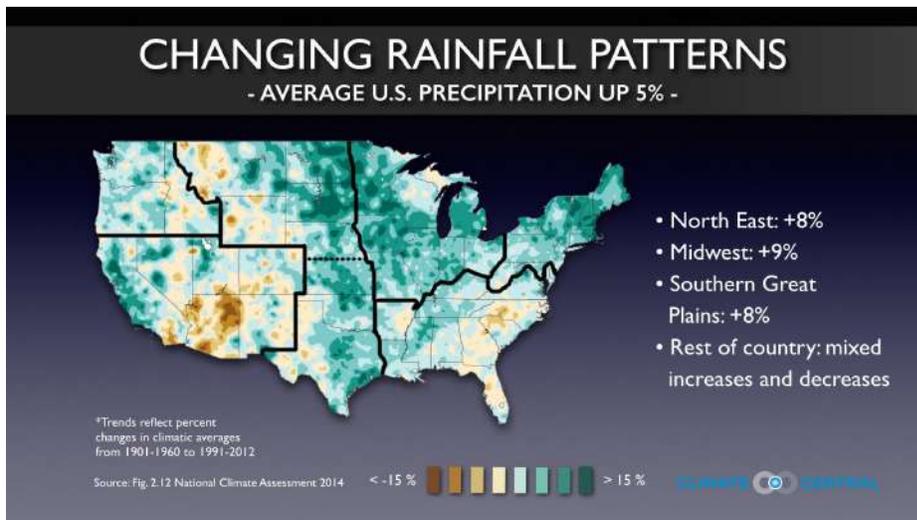
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Our Haulers

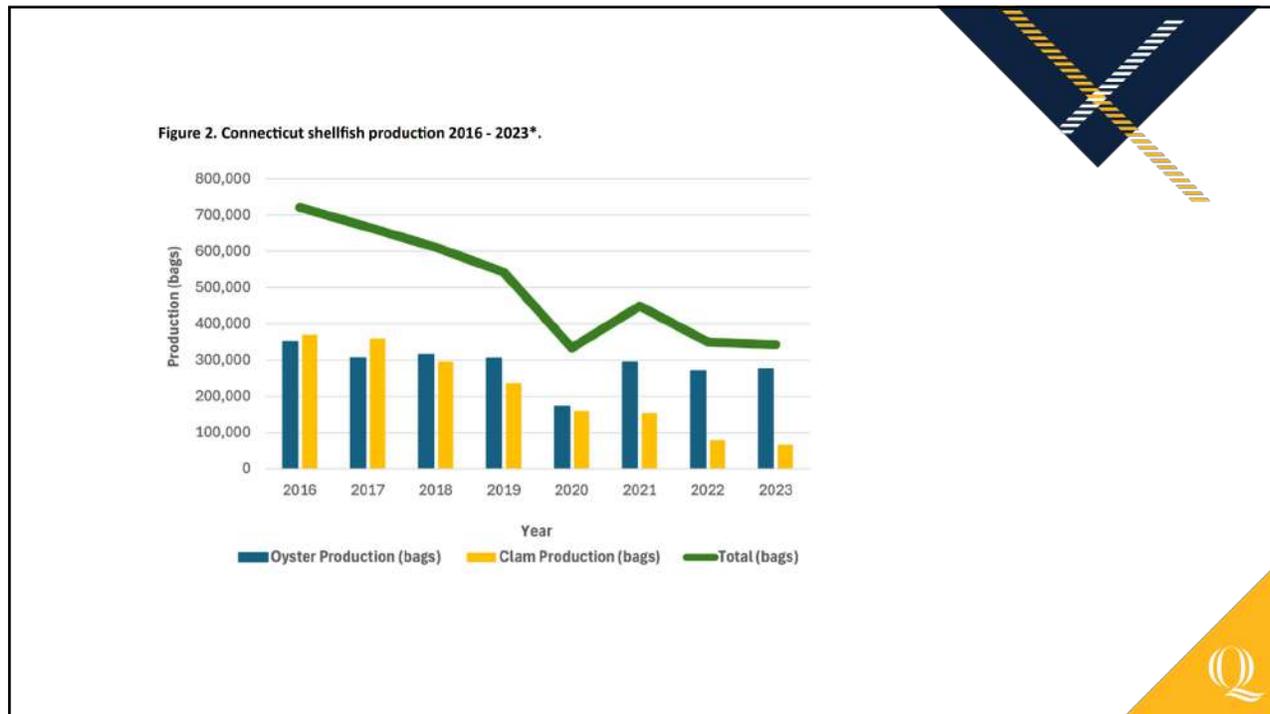


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Why is this work important?



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Conclusions & Future Directions

- Continue sampling the QU River through late Fall and Winter months
- Analyze river samples for total coliform, E.coli and other water quality parameters such as pH and conductivity
- Compile seasonal data for trends at each location, including rainfall
- Continue to monitor the health of the Quinnipiac River (Seasonal)
 - Plasticizers, Phthalates etc. → Source
- Developing a method for PFAS detection
- Continue to survey for Chinese mitten crab




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Acknowledgments

- Quinnipiac University College of Arts and Sciences
 - Undergraduate & Graduate Students
- QU River Fund, managed by the Community Foundation for Greater New Haven (2015-present)
- Harry M. Pylypiw, Ph.D.
- Nancy Alderman



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